

An Algorithm for the Automatic Analysis of Signals From an Oyster Heart Rate Sensor

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Abstract—An *in situ* optical oyster heart rate sensor generates signals requiring frequency estimation with properties different to human ECG and speech signals. We discuss the method of signal generation and highlight a number of these signal properties. An optimal heart rate estimation approach was identified by application of a variety of frequency estimation techniques and comparing results to manually acquired values. Although a machine learning approach achieved the best performance, accurately estimating 96.8% of the heart rates correctly, a median filtered autocorrelation approach achieved 93.7% with significantly less computational requirement. A method for estimating heart rate variation is also presented.

Index Terms—Biomedical signal processing, frequency estimation, machine learning.

I. INTRODUCTION

HEART rate and heart rate variability in intertidal invertebrates are proxies for physiological responses (such as oxygen transport, neural feedback, stress) and tracking these quantities is highly informative. For example they allow the impact of external environmental factors to be quantified and enable optimization of conditions for breeding and consumption stock. Consequently there is a need for heart rate monitoring systems for these animals.

Measurement of heart rate in humans is a well established and mature science [1]; ECG systems detect electrical signals generated by the heart. These signals are amenable to analysis and were first analysed by computer in the 1960s [2]. Real-time heart rate computer analysis systems were in use by the 1980s [3]. Analysis is based on detecting the repetition and shape of the ECG signal structure referred to as the QRS complex. Unfortunately such approaches are not applicable to intertidal invertebrates (such as the oyster) which possess a shell preventing use of externally applied

ECG sensors. Early experiments on the bivalve class of intertidal invertebrates observed heart rate by measuring the heart's electrical impedance via implanted electrodes [4]. However, it is critical that the system measuring heart rate does not modify the animal's heart rate by (for example) increasing animal stress. To mitigate the invasive nature and degradation of electrical contacts in the impedance approach an alternative optical coupling method was proposed [5]. This approach illuminates the heart with an optical signal and detects the intensity of reflected light which is modified as a consequence of any motion of the heart's surface. For thin shelled intertidal invertebrates a near infrared source and detector are collocated and positioned on the shell's outer surface. The NIR signal propagates through the shell without need for drilling into the shell. For thick shelled invertebrates such as the oyster, the approach is similar with the exception of drilling a small hole into the shell [6] which would otherwise block the optical signal. The light emitter and detector are mounted and sealed to ensure the integrity of the shell is maintained. Optical techniques avoid direct contact with the animal; however, the approach introduces its own problems in terms of signal analysis due to signal complexity as pointed out in a recent review paper [7]. Therefore there is a need to develop robust signal analysis approaches operating over long installations possible with these sensors. In this paper we analyze the signal from a sensor *in situ*, and develop algorithms to detect short term average heart rate and variation of heart rate within short (20s) signal sequences.

The oyster biosensor [8] (Fig. 1) provides information on the animal through both shell gape and heart rate detection. Gape refers to the opening of the two shells which occurs when the oyster is submerged and feeding and/or taking in oxygen. The gape sensor (based on a Hall effect sensor) detects open and closed states of the oyster shell by measuring changes in the output voltage of a transducer located on the upper shell in response to a magnetic field induced by a magnet located on the lower shell. The heart rate sensor illuminates the oyster heart with a fixed intensity 950nm (NIR) optical signal generated by an LED source, and measures the reflected optical intensity via an IR diode detector. The LED source and detector are collocated and mounted within 5mm of the heart surface. The sensor responds with a time constant of 20 μ s to reflections within a cone angle 55 degrees from the sensor axis out to a range of 5mm. The principal of operation of the heart rate sensor is that the heart's surface reflects light which is measured by the IR sensor. Any motion

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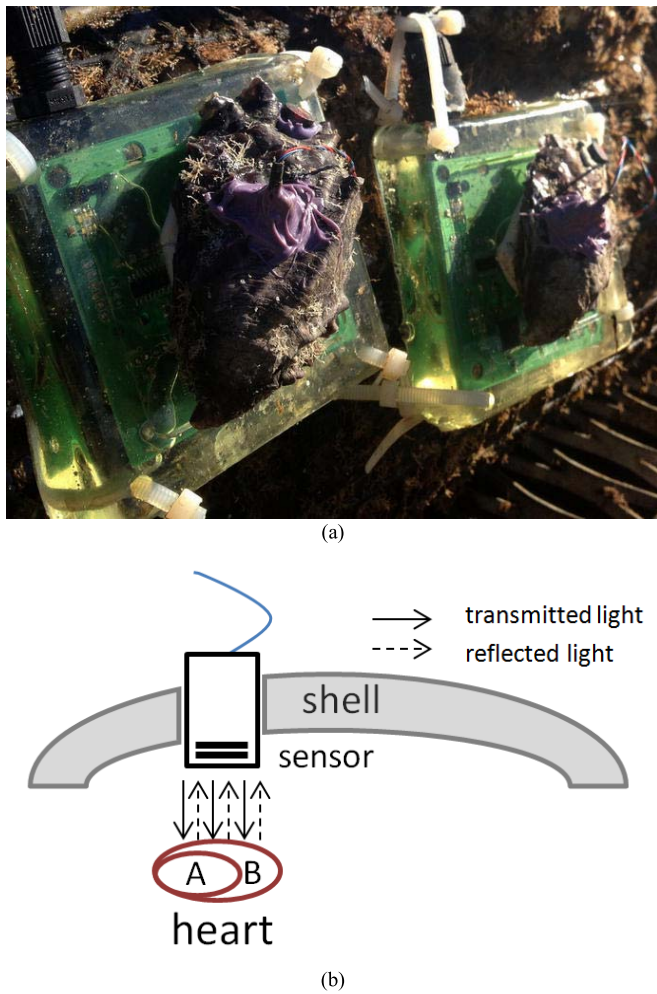


Fig. 1. Oyster attached to biosensor. (a) Photo of sensor and oyster. (b) Schematic of sensor inserted into hole in oyster’s shell. Heart size depends on whether it is in A: contraction phase (systole) or B: expansion phase (diastole). Reflected light rays shown for expanded heart. For contracted heart fewer rays will strike heart with corresponding change in detected signal level.

in the heart causes perturbation in the surface geometry, thus modifying the intensity of the reflected optical signal. If the heart’s surface geometry varies periodically (as is the case for a beating heart) the measured signal exhibits a period equal to that of the heart’s motion. Measurement of the signal period is used to infer heartbeat period and consequently heart rate. The sensor is aligned such that signals are detectable both when the oyster is open and closed. Reflected light intensity is sampled at 100Hz over 20 seconds, resulting in sequences containing 2000 samples. This sampling regime is selected to capture multiple heartbeats per sequence, which are expected to have durations within the range of 0.5 – 5 seconds [9]. Currently one sequence is recorded approximately every six minutes for the duration of sensor installation, which has been running for nine months. One gape value is recorded for each sequence. Three classes of sequence occur: saturated (Fig. 2a), aperiodic (Fig. 2b), and periodic (Fig. 3). Any algorithm must classify a sequence as one of these three classes, and determine the heart rate and heart rate variation of sequences classified as periodic.

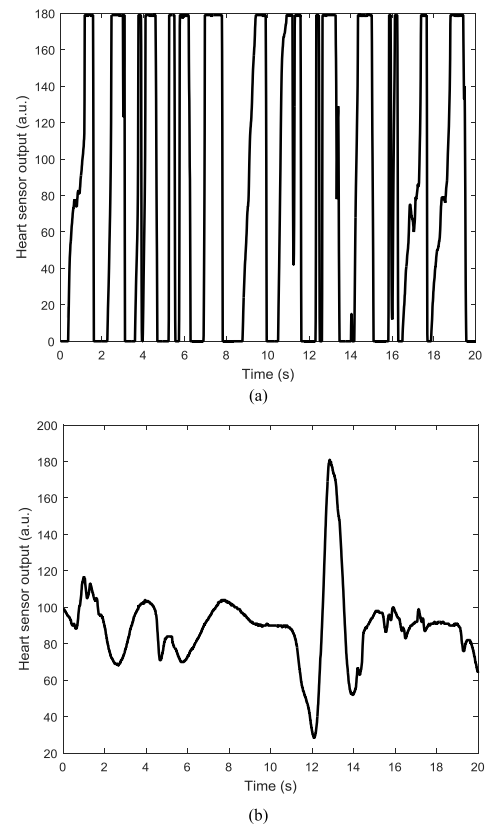


Fig. 2. Examples of saturated (a) and aperiodic (b) sequences.

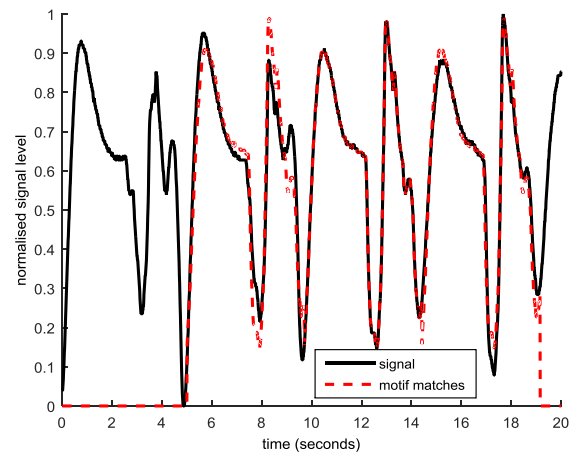


Fig. 3. Example of periodic time domain signals including 4 heart beats. Note peak value within beat occurs for first pulse in two beats up to 10s, and second pulse for last 2 beats. Sequence motif (described in Section II) shown in red/dashed overlays signal from 9.5 to 14.5 seconds. Best shifted motif matches shown from 5.0-9.5 seconds and 14.5-19 seconds. 5-9.5 and 15-20 seconds.

Saturated signals likely occur due to the presence of external light sources, or due to sub-optimal biasing of detector electronics. Aperiodic signals likely occur in the absence of strong signal coupled off the heart surface, or absence of a beating heart. Periodic sequences result as a consequence of the beating heart periodically modifying the intensity of the reflected signal coupled into the detector. The dual chamber heart comprises a smaller ventricle and larger

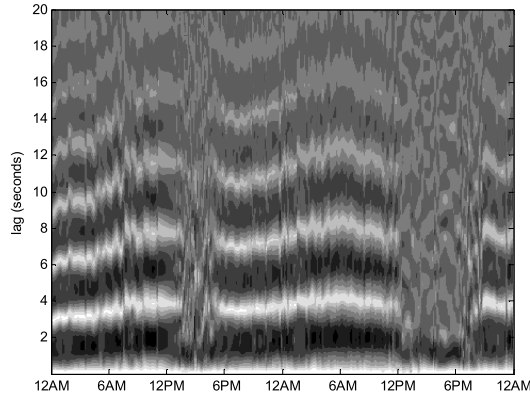


Fig. 5. Lag domain amplitude (white positive, black negative) from sequences gathered over a 48 hour time frame November 3–November 4, 2014.

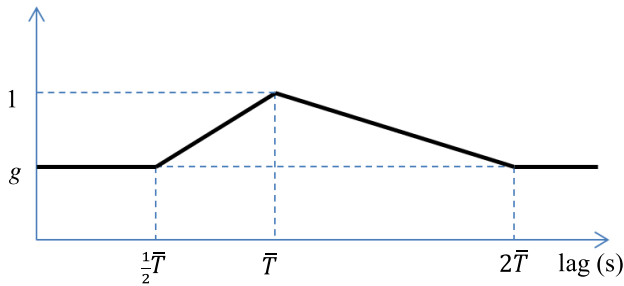


Fig. 6. Tracking estimator's weighting function for scaling autocorrelation approaches' amplitude in the lag domain.

dual chamber beating is not erroneously selected. Alternative methods such as the NAC and HWE exist to compensate for this.

Despite the gaps between sequences, the slow change in oyster heart rate results in similar periodic structure which is apparent when plotting the autocorrelation function for successive samples (Fig. 5). This long term stability and the HWE lag domain weighting approach motivated a simple tracking (TR) estimator which is biased towards selecting heart rates similar to those estimated from the previous sequences where N_{TR} is the number of sequences used in the calculation. The motivation is to reduce the occurrence of erroneous heart rate estimates at half and double the actual rate (typical errors for estimators on signals with periodic beats containing double peak structure). To achieve this bias the lag space is weighted with a piece-wise linear window. The window was maximal (and unit valued) at the mean detected period (\bar{T}) over the previous N_{TR} sequences, and linearly decreases for smaller (and larger) lags until reaching a minimum value $g < 1$ at lags corresponding to half (and double) mean periods. All lags outside this range are also weighted by g , the weighting window is shown in Fig. 6. Optimal values of g and N_{TR} were estimated for the tracking estimator.

We also applied a median filtering (MF) estimator applied to the best performing basic estimator:

- 1) *The median period was calculated for the N_{MF}^1 sequences prior to and including the sequence being analysed.*

- 2) *Sequences with periods more than 20% different to the median period were removed from the set of N_{MF}^1 periods.*
- 3) *From remaining sequences the median period was calculated for the N_{MF}^2 sequences prior to and including the sequence being analysed.*

The longer time frame corresponding to N_{MF}^1 provides robust longer term averages, whereas the potentially shorter time frame N_{MF}^2 allow accurate estimations of short term transitions in heart rate. These values were optimally evaluated for the median filter approach.

Finally we implemented a machine learning approach using nine frequency estimators (within dashed block in Fig. 4) as input features. Three WEKA [19] implementations were assessed including a three layer multilayer perceptron (MLP) with six hidden nodes (larger networks showed no improvement in performance). A Linear regression (LR) and a support vector regression (SMO).

Performance of the techniques was quantified by comparing estimated heart rates with values acquired by manually observing sequence class and heart rate within the first 2 months of collected data. After removing saturated sequences from the data set, sequences were randomly selected and classified as either periodic or aperiodic based on visual inspection until 100 sequences were labelled for each class (200 in total). Sixteen regions of consecutive periodic signals were identified and the periods of 9 sequences (evenly spaced throughout each region) were measured by visual inspection resulting in a total of 144 measurements. Ground truth periods were calculated by linearly interpolating the measured sequence periods across each span of sequences resulting in a total of 1450 sequences with ground truth period data. For the machine learning approach the 1450 sequences were split into 10 sets of 145 consecutive sequences and a 10-fold cross-validation used to evaluate performance, training on 9 sets and testing on the remaining set.

In addition to heart rate the variation of heart rate is also of interest as an indicator. Whereas heart rate variation over longer time frames can be calculated as the difference between heart rates estimated from sequences with the desired temporal separation, short term (intra-sequence) variation requires an estimate of the duration of each heartbeat. Because the structure of the heartbeat signal can change within a sequence a motif approach was used to estimate the temporal locations of each beat within the sequence. For each sequence a subsequence of length T was identified as the motif that is most representative of the multiple beat signals within the sequence. Temporal shifts of the motif within the sequence that best reconstructed the entire sequence were used to estimate beat delay and consequently estimate heart rate variance.

A subsequence $S_{t_0}^T(t)$ of sequence $S(t)$ is uniquely defined by length T and starting time t_0 such that $S_{t_0}^T(t) = S(t + t_0)$. We zero pad the sequence to length L such that $S_{t_0}^T(t) = 0$ for $t > T$. The sequence motif is the subsequence that captures more of the periodic structure in the signal than any other subsequence. To quantify this we calculate the

cross-correlation $C(\tau)$ of $S_{t_0}^T$ with S :

$$C(\tau) = \mathcal{F}^{-1}\{\mathcal{F}\{S_{t_0}^T(t)\} \cdot \mathcal{F}^*\{S(t)\}\} \quad (2)$$

Where \mathcal{F} is the Fourier operator and $*$ the complex conjugate. The cross-correlation requires normalization to eliminate bias towards selecting motifs in the high amplitude regions of the signal. Both the power in the sub-sequence and the distribution of power throughout the sequence contribute to this bias. A normalization factor is calculated from the cross correlation of instantaneous signal power S^2 with a zero padded step function H of duration T :

$$N(\tau) = \mathcal{F}^{-1}\{\mathcal{F}\{S^2(t)\} \cdot \mathcal{F}^*\{H(t)\}\}$$

Where $H(t) = 1$ for $t \leq T$ and $H(t) = 0$ for $t > T$. Consequently $N(t_0)$ is the power in the subsequence and $N(\tau)$ the power in the target sequence starting at time τ of length T .

A subsequence $S_{t_0}^T$ can repeat L/T times throughout the sequence of length L . We evaluate the peaks of the cross-correlation at temporal lag steps of size T corresponding to $\tau_i = iT + t_0 \text{ modulo } T$, where $i \in \{1, \text{floor}(L/T)\}$. To calculate heart rate variation we allow for period drift within a sequence and therefore look for maximum values within a range $\{\tau_i - kT, \tau_i + kT\}$ where k depends on the expected maximum heart rate variation within a sequence. Finally given the estimated heart rate L the sequence motif is defined by delay t_m which corresponds to the subsequence delay t_0 which maximizes the following expression:

$$t_m = \underset{t_0}{\operatorname{argmax}} \left\{ \sum_{i=1}^{\text{floor}(L/T)} \max_{\tau \in \{\tau_i - kT, \tau_i + kT\}} \left[\frac{C(\tau)}{N(t_0)N(\tau)} \right] \right\} \quad (3)$$

The expected maximum heart rate variation within a sequence was set to $k = 0.1$. After detecting the best motif the temporal shifts of the motif τ that resulted in maximizing expression (3) are used for estimating beat to beat variation estimation within a sequence.

III. RESULTS

The first 2 months of available data was assessed. Of the 9000 sequences, approximately 14% were saturated, 61% periodic, and the remainder aperiodic. Of the approaches the ACM generated the best classification of the aperiodic/periodic signals achieving 89% accuracy with a detection threshold of 1/3 with respect to the 200 sequences for which periodic/aperiodic ground truth was available. Therefore ACM was used for the classification stage in Fig. 4.

In terms of the saturated signals, a histogram of saturated signals against time of day is shown in Fig. 7 and indicates this detection approach is heavily influenced by the presence of sunlight. An explanation for the generation of saturated sequences (Fig 2a) in the presence of sunlight is that the IR sensor is detecting bursts of sunlight. The presence of waves on the water surface directly above the oyster results in an instantaneous distribution of sunlight below the water surface. This distribution contains regions of increased sunlight due to a focusing effect via refraction through the

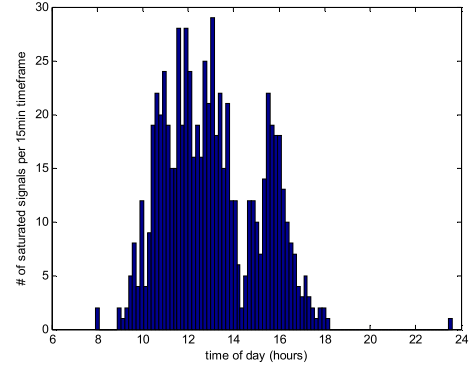


Fig. 7. Histogram of saturated sequence class against time of day.

TABLE I

ACCURACY RESULTS FOR IMPLEMENTED METHODS. *TR USES ACM, $g = 0.6$, N_{TR} CORRESPONDS TO 4.5 HOURS. **MF USES ACM, N_{MF}^1 AND N_{MF}^2 CORRESPOND TO 4.5 HOURS AND 1 HOUR, RESPECTIVELY

Approach		HR estimates % accurate	Heart beat period estimates % split into low (L), correct (C), and high (H) ranges		
			L	C	H
Lag	ACM	83.2	0.3	89.7	10.0
	CCA	82.7	0.1	88.8	11.1
	NAC	11.6	2.6	11.9	85.5
	HWW	71.5	1.9	77.9	20.1
Time	AMD	13.5	0.0	14.0	86.0
	MLH	7.0	0.0	7.0	93.0
Frequency	CEP	33.0	3.7	35.7	60.6
Aggregated	TR*	85.2	0.3	91.3	8.4
	MF**	93.7	0.0	99.9	0.1
Machine Learning	MLP	96.3	0.7	99.2	0.1
	LR	96.7	0.7	99.2	0.1
	SMO	96.8	0.7	99.2	0.1

air/water interface. Wave motion then results in temporal variation of sunlight incident onto the oyster and consequently coupled into the detector.

The ultimate aim of heart rate estimation is to find associations with physiological response of the oysters. For analysis by biological scientists a 90% accuracy was targeted. Furthermore due to the interpolated nature of the generated ground truth values, the ground truths were estimated to be within 4% of the actual periods. This was based on comparing accuracy from a coarser interpolation. Consequently a heart rate estimate was deemed accurate if it was within 10% of the manually acquired ground truth. The percentages of heart rates estimated accurately for each approach are shown in Table I. The most accurate methods are the machine learning approaches followed by the aggregated approaches. The ACM achieves the highest accuracy of the single (lag, time and frequency) estimators (83.2%). Multiple ACM heart rate estimates were then aggregated to calculate

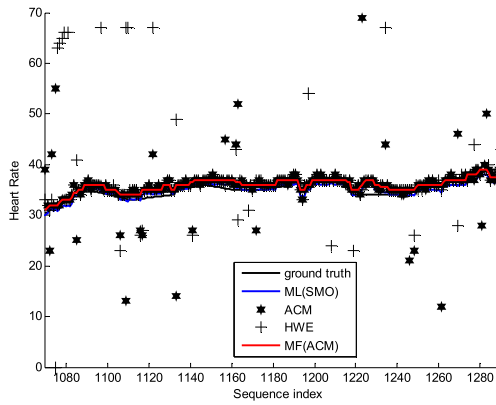


Fig. 8. Plot of estimated heart rates for a selection of approaches.

the Tracking and Median Filtered estimators. Optimal values of N_{TR} corresponded to time frames of 4-5 hours with optimal $g = 0.6$ giving an accuracy of 85.2%. The median filter approach achieved 93.7% with values of N_{TR} and N_{TR} corresponding to 4.5 hours and 1 hour respectively. Note these optimal values are sensitive to the level of heart rate variation over time frames corresponding to hours. Values of N_{TR} and N_{TR} corresponding to 2 hours and 20 minutes respectively still achieved 92% accuracy, but may be more accurate for estimating heart rates where variation is more rapid than the current installation. Finally the Machine learning approach achieved superior results, the best approach (based on a support vector regression) achieving 96.8% accuracy.

The accuracy of any approach is dependent on two factors: the ability to estimate the period within the sequence accurately, and the ability to avoid erroneously selecting double and half periods. Each estimate of heartbeat period was compared to the ground truth. If the estimated period was within the range $2/3$ to $3/2$ multiples of the ground truth period the estimate was categorized as being in the correct range. For period estimates below and above this range, the estimate was categorized as being in the low or high range respectively. The percentages of estimated periods categorized into these ranges by each approach are shown in Table I. For lag based approaches the accuracy is strongly related to the erroneous selection of longer periods. The bias of ACM and CCA to lower periods via lag domain weighting resulting from sequence zero padding is evident, whereas the normalised approach which removes this weighting suffered the most. The two pulse structure of the heart rate signals has the potential to generate erroneous half period peaks in the lag domain. Despite the lower lag bias of the ACM and CCA the ACM and CCA were not susceptible to the half period error. The time domain methods exhibited similar performance to the NAC. The CEP had the highest of estimates in the low period range, as a frequency domain method this corresponds to the CEP selecting the second harmonic, resulting in half period estimates. The MF approach was highly successful at avoiding half and double period estimates; the averaging effect over multiple samples did not dramatically reduce accuracy. Information in the multiple estimators was utilized by the machine learning approach to improve the accuracy beyond the median filtered results.

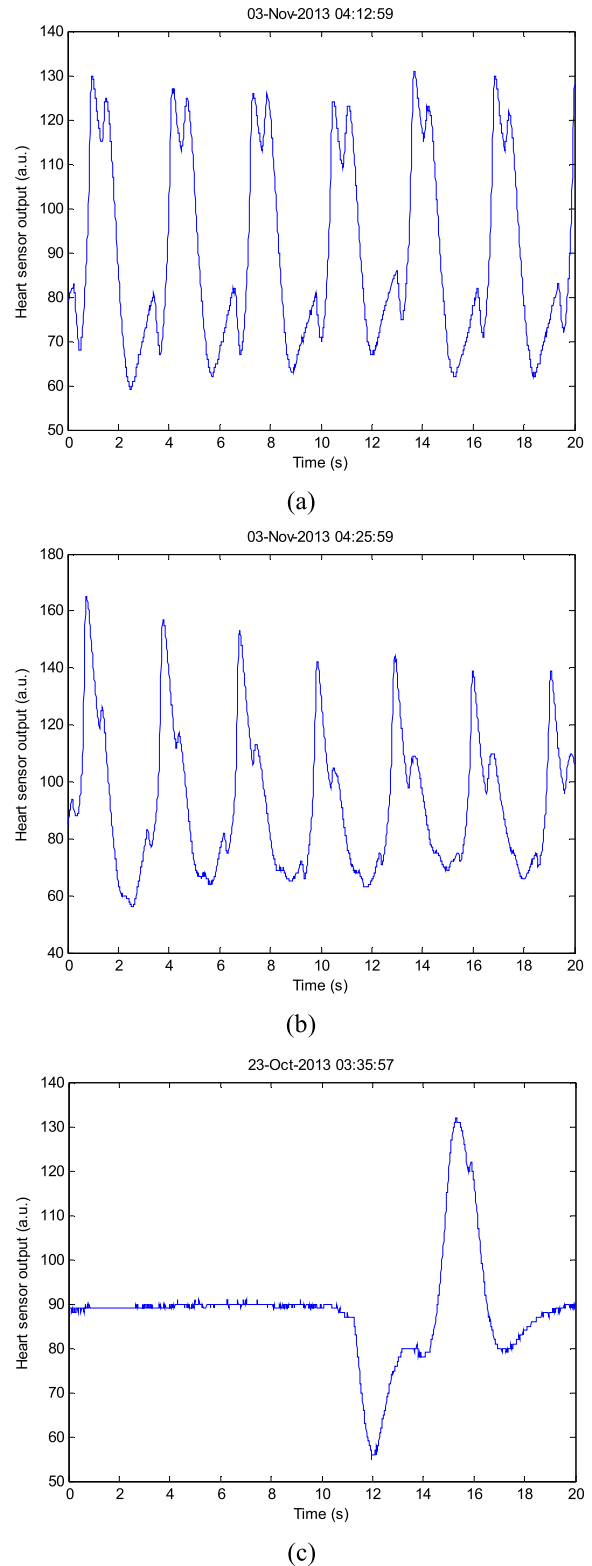


Fig. 9. (a)-(b) Two sequences of 20 s duration from early region of Fig. 3 timeframe at ~4:13AM and 4:25AM (13 minutes apart). (c) Example sequence including single “beat.”

The results show a wide variation in the ability of the methods to determine heart rate which is strongly influenced by the bias of the various methods towards detection of heart rates at double and half the correct heart rate. An example range of sequences is shown in Fig. 8 along with estimated

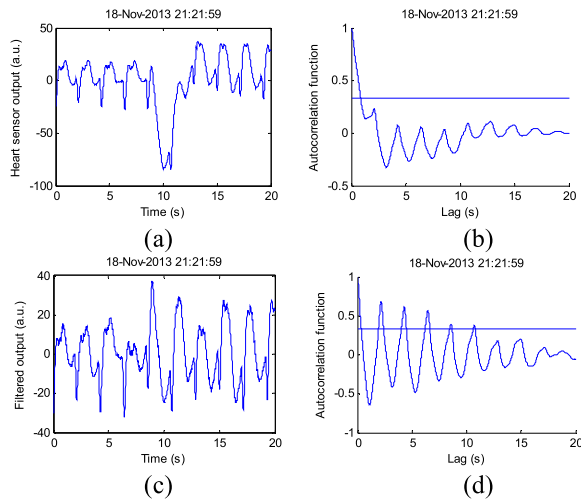


Fig. 10. (a) Original sequence (DC removed). (b) Lag domain of sequence. (c) 2 Hz high-pass filtered sequence. (d) Lag domain of filtered sequence.

heart rates; the ML and MF closely follow the ground truth, whereas ACM and HWE exhibit erroneous readings on some sequences.

For heart rate sensors deployed on cheap, lightweight hardware the MF(ACM) approach is ideal as the more accurate machine learning approach requires labelled training data and additional computational complexity. However; for offline analysis with labelled data the machine learning approach provides the most accurate results.

The signals exhibit a number of unique properties. For example the signals can switch between regions of periodic and aperiodic signals (observable in lag domain Fig. 5). Although the heart rate varies slowly the signal shape can vary rapidly. This is demonstrated by the sequences in Fig. 9a, b which are separated by approximately 13 minutes. These sequences are periodic with identical heart rates but exhibit significantly different shapes. Sudden signal deviations often occur either in isolation (Fig. 9c) or within a periodic sequence (Fig. 10a). Isolated deviations likely indicate the presence of a short term geometry variation caused either by a single heartbeat or the shell opening and closing. Higher frequency sampling of gape would identify the cause. Deviations within periodic sequences are often ~ 2 s in duration, with a corresponding change in the periodic structure either side of the deviation. An example of this is seen by comparing the signal waveform in Fig. 10a 0s-5s versus the 15s-20s signal waveform. We believe these dips occur when the oyster rapidly closes and opens the shell, thus changing the coupling geometry. Furthermore these dips cause problems for the ACM due to the first peak (2s in Fig. 10b) becoming obscured by the spread of the self coherence peak (0s in Fig. 10b). The spread is introduced as a consequence of the sequence mean value being significantly different to the periodic structure local mean value resulting in temporarily localized DC offsets which extend the coherence time. The two sequences recorded adjacent to Fig. 9a were correctly identified with 2s period. One solution is to high pass filter the sequence which removes the localized DC offset and exposes

the lag peaks (Fig. 10d). However this lowers the sensitivity of the method to heart rate detection in other sequences. As these errors are present within single sequences, the median filtering approach eliminates these errors.

IV. CONCLUSION

We discussed a range of properties of signals generated by a novel oyster heart rate monitor and consequences on signal analysis. These properties include sudden changes in signal level, transition from periodic to aperiodic signals, large saturated signal interference, and evolution of the periodic structure within the sequences. We implemented a range of existing methods for estimating heart rate, along with some tailored approaches aggregating multiple sequences, and a machine learning approach. The approaches were all able to estimate heart rate. The lag domain approaches biased to shorter lags were the best single sequence simple estimators achieving 83.2% accuracy. However none of the existing methods achieved better than 90% accuracy. This accuracy required a machine learning approach which achieved the best results (96.8%) using all available estimators as input features. Alternatively an approach utilizing multiple consecutive sequences (the median filtered autocorrelation method) was required which achieved acceptable accuracy (93.7%). The ultimate choice is determined by available computational resources. Future work would require implementation of the algorithms within the sensor platform and require computationally efficient means in calculating heart rate variation.

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Sarah Andrewartha received the Ph.D. (Hons.) degree from Melbourne's La Trobe University. She focused on understanding how the environment influenced the development and physiology of a broad range of animals. She was a Post-Doctoral Fellow with the University of North Texas, where she was involved in research on the development of acid-base regulation in chicken embryos, and epigenetic inheritance patterns in response to changes in the environment of *Daphnia* (tiny freshwater crustaceans with a transparent body, often used in biological research). She is currently a Comparative Physiologist and Post-Doctoral Fellow with the Commonwealth Scientific and Industrial Research Organisation, Hobart, Australia. She uses biosensors to monitor the effect of environmental changes on shellfish and fish physiology. In the future, this information will be used by aquaculture farmers to assist farm management, based upon measurements of animal fitness. She is examining body temperature influences on metabolism, ventilation, and blood acid-base regulation in developing Tamar wallaby joeys, and the consequences of the nest environment for young reptiles and amphibians.



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