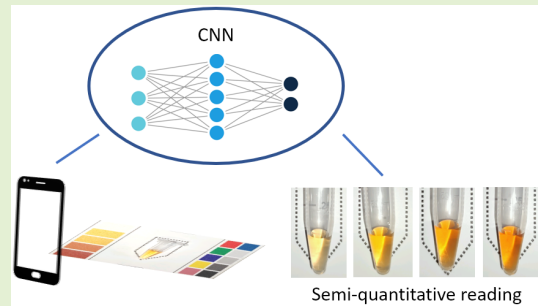


Semiquantitative Determination of Thiocyanate in Saliva Through Colorimetric Assays: Design of CNN Architecture via Input-Aware NAS

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Abstract—This article presents a novel method enabling point-of-care (POC) testing of thiocyanate concentration in saliva. Thiocyanate is an important biological marker; its levels are linked with diseases such as cancer and neurodegeneration. Hence, monitoring this marker frequently can positively impact users' lives. In the proposed setup, the goal is a semiquantitative reading of thiocyanate concentration from colorimetric assays in solution; the user-friendly, yet accurate readout procedure relies on a smartphone camera and is designed to be robust against moderate changes in indoor lighting conditions. The readout procedure exploits the capabilities of convolutional neural networks (CNNs) to fully profit from a setup involving a custom color chart and the assay vial. Thus, a data-driven strategy is adopted to deal with color distortions caused both by lighting conditions and by postprocessing operations embedded in the smartphone camera. A neural architecture search (NAS) procedure explicitly tuned for the problem at hand drove the design of the custom CNN architecture. The method has been tested using a collection of real-world data and compared with existing approaches. The results presented in this article show an increase in accuracy up to about 14% with respect to state-of-the-art methods.



Index Terms—Colorimetric test, convolutional neural networks (CNNs), neural architecture search (NAS), point-of-care (POC), saliva, smart sensor.

I. INTRODUCTION

A BIOLOGICAL marker (*biomarker*) is “any substance, structure, or process that can be measured in the body or its products and influence or predict the incidence of outcome or disease” [1]. Thiocyanate is an important biomarker that strongly correlates with a person’s well-being: its levels are linked with diseases such as certain types of cancers, atherosclerosis, and neurodegeneration [2], [3], [4], [5], [6] as well as with bacterial infections and tobacco smoke expo-

sure [7], [8], [9], [10]. Monitoring this biomarker frequently can positively impact users’ lives, but many technological challenges have to be overcome to develop a fast and easy-to-use point-of-care (POC) sensing device. In fact, although thiocyanate can be sampled noninvasively from saliva, most techniques to assess its concentration require special instrumentation and knowledge, raising issues about costs, assay portability, or ambient conditions stability in the case of instrument-free devices [11].

This article focuses on the semiquantitative reading of thiocyanate concentration in saliva from colorimetric assays in solution [7]. The target is to develop an automatic readout procedure that is robust against moderate changes in indoor lighting conditions. The adopted setup relies on a smartphone camera and a custom color chart that appears beyond the assay vial when the picture is grabbed in indoor settings. Such an approach may lead to an easy-to-use POC device: the user takes a picture of a test with the smartphone and then automatically receives the result on the smartphone display. It is known from the literature that smartphones can serve as cheap and portable readout systems for POC design [12], [13], [14], [15]; this, in turn, leads to inexpensive and fast solutions, which can be used even by untrained users. However, the interpretation of colorimetric test kits is challenging, even in

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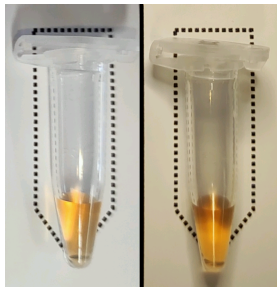


Fig. 1. Same test vial captured under different ambient light conditions. The different illumination strongly impacts the color grade and uniformity as well as the formation of unpredictable shade patterns.

the case of commercial products [16], [17], [18]. This process may lead to errors and low accuracy, practically limiting the assays only to those exhibiting strong and distinguishable colors [19]. Hence, the availability of reliable readout procedures can further enhance the features of smartphones as sensing systems.

Recent reviews [20], [21] highlight the lack of robust solutions for the readout of colorimetric analysis in ambient light, as severe limitations affect sensing methods based on smartphone cameras. These issues, in turn, prevent accurate quantitative and semiquantitative colorimetric analyses. Two main issues should be mentioned. First, lighting conditions impact the image captured by the camera. Second, unavoidable low-level, postprocessing operations embedded in the smartphone camera further increase color distortions. Furthermore, assays in solution represent a major problem as the shape and the texture of the vials may produce color gradients, depending on the angle from which the illumination hits the test vial (see Fig. 1).

The main contribution of this article lies in a novel approach based on convolutional neural networks (CNNs) for automated readout of colorimetric assays in solution under ambient light conditions. The proposed approach exploits the effectiveness of CNNs at solving nonlinear problems to fully profit from a setup that involves a custom color chart (Fig. 2) along with the assay vial. Accordingly, the CNN-based model takes advantage of the presence of reference color patches in the captured image to manage color distortions caused both by lighting conditions and postprocessing operations embedded in the smartphone camera. In practice, a data-driven strategy is adopted to bypass explicit color correction techniques, which lack robustness.

A neural architecture search (NAS) procedure automatically generated and evaluated a set of candidate neural network architectures. Such an approach led to a custom CNN designed to tackle the problem at hand: the NAS procedure was specifically designed to select an input-aware architecture, as the candidate networks also differed in the relative weight assigned to each reference color included in the custom chart.

The contribution of the article can be summarized as follows.

- 1) An automatic readout procedure designed for smartphones that can assess thiocyanate levels in saliva from an assay performed in solution in ambient light.

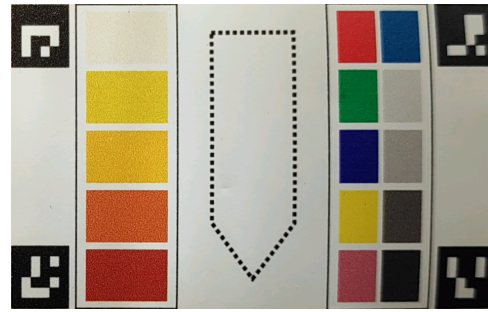


Fig. 2. Custom chart with five reference color patches (on the left of the vial-shaped outline) and ten color correction patches (on the right). The four ArUco markers can be seen on the corners.

- 2) A data-driven procedure to automatically generate and evaluate neural network architectures and optimize the input representation.
- 3) A NAS search space suitable for colorimetric diagnostic tests performed in solution using smartphone cameras and paper support in ambient light.

Experimental results on real data confirmed the reliability of the proposed approach. The custom CNN architecture yielded from the NAS outperformed state-of-the-art solutions targeting similar problems [22], [23], showing up to a 14.8% increment in accuracy on a ten-class classification problem.

II. RELATED WORKS

POC devices have recently attracted a surge in attention in various branches of research and clinical diagnostics [24], [25], [26]. Several types of POCs have been developed for different applications [27], [28], [29], [30] including lab-on-a-chip [31], lab-on-a-disk [32], microfluidic paper-based analytical [33], [34], lateral flow devices [35], [36], and isothermal nucleic acid amplification [37].

Smartphones can effectively support the design of smart POC devices [12], [13], [14], [15]. More specifically, digital cameras [20], [21] play a major role in low-cost sensing as an alternative to spectrophotometric analysis [38], [39]. This, however, is a challenging task mainly due to the instability of the color captured. Existing solutions can be grouped based on physical setup and algorithm.

In some works, enclosures and supports [40], [41] eliminate the detrimental effect on the color of ambient light, using the smartphone flashlight or internal fixed lights as the only illumination source [42], [43]. The presence of external hardware makes these approaches less commercially appealing than operations in ambient light.

Other methods use color correction techniques from computer vision [44], [45]: the setup involves a color board placed close to the assay. A color correction matrix (CCM) maps each pixel to its “true” color. CCM can be obtained using different algorithms: linear color correction, polynomial color correction, and root-polynomial color correction. As stated in [46] and [47], the latter provides the best and most consistent results. However, these methods require the color board and the test to be hit by the same, constant lighting. Moreover, the individual colors of the board should not be saturated, which makes them prone to user errors.

Machine learning (ML) can learn the interaction between light and the assay automatically from data. Most ML methods deal with the fluctuations of ambient light by using reference color charts, which can be put close [48], [49] or farther [22] from the assay. These colors are similar to those appearing in the test and are expected to be affected by light conditions coherently. Flaucher et al. [22] used three different approaches to compare the test and ground-truth colors: Hue value comparison, matching factor, and Euclidean distance, with the last two algorithms computed in the HSV color space.

Among ML techniques, DNNs excel in image processing [23], [50], [51]. In [23], a CNN classified albumin concentration values in urine from a paper-based assay using different smartphones' cameras, evaluating also the performance improvement gained using the smartphones' flash. This method, however, does not provide any additional information to the neural network about the lighting in which the test is taken, with an impact on the overall accuracy.

III. METHODOLOGY

This section presents the method for semiquantitative reading of thiocyanate concentration in saliva using colorimetric assays in solution. The research achieves robustness even in indoor ambient light conditions, where the direction, strength, and color of the lighting may vary significantly.

The approach uses fixed reference colors put close to the assay vial that allows the CNN to estimate the light's properties. A NAS procedure selects automatically a suitable CNN architecture in combination with a set of reference colors.

In the following, Section III-A presents the method, while Section III-B introduces the NAS procedure.

A. CNN Architecture for Semiquantitative Reading of Salivary Thiocyanate Concentration in Ambient Light

Our goal is to read the thiocyanate concentration present in saliva, indicated as T , in (1), from a colorimetric assay in solution. The underlying hypothesis is that the assay reflectance R is related to T [7]

$$T = g(R). \quad (1)$$

The smartphone camera provides information about R . In general, cameras perceive colors as a function of the spectrum of the source of light L which strikes a certain dark object and its surface reflectance. Given the same reflectance for an object, a sensor perceives its color differently depending on the spectrum of the light hitting it and its sensitivity S . In our case, the liquid state of the assay is an issue, as the perceived color is altered by the direction from where the light hits the vial and by its strength (see Fig. 1).

As a result, the assay color perceived by the camera C_P is

$$C_P = f(R, L, S). \quad (2)$$

Equation (2) clarifies that to properly assess R , one should know L and S in addition to C_P . Accordingly, estimating T requires solving the inverse problem

$$T = g(f^{-1}(C_P, L, S)). \quad (3)$$

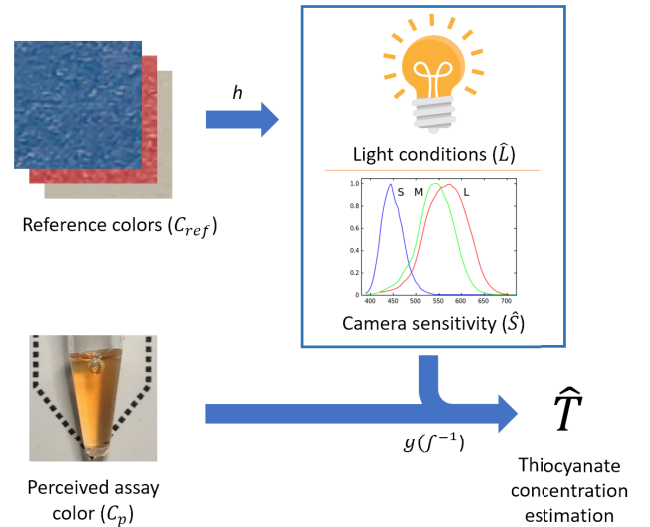


Fig. 3. Theoretical passages needed to retrieve the thiocyanate concentration value from the data obtainable through a picture using our reference color card.

However, both L and S are unknown. The present approach tackles this issue by estimating S and L . Reference colors are placed close to the salivary test by adopting the custom chart of Fig. 2. The chart contains 15 color patches: ten patches have been extracted from popular color correction charts; the remaining five set a reference color scale for the assay with different thiocyanate concentrations. The card contains four ArUco markers [52] on its corners (as per Fig. 2); such markers facilitate the alignment process and the correction process for skewed images, reducing the impact of phone tilt and rotation.

The overall goal is to estimate S and L by using the 15 colors $\{C_{ref,i}; i = 1, \dots, 15\}$ captured by the camera. Fig. 3 schematizes the setup. A function h , shown in (4), estimates S and L using the true reflectance value of the reference colors (upper part of Fig. 3)

$$\hat{S}, \hat{L} = h(\{C_{ref}\}). \quad (4)$$

Then, \hat{L} and \hat{S} are exploited in the solution of the inverse problem (3)

$$\hat{T} = g(f^{-1}(C_P, \hat{L}, \hat{S})). \quad (5)$$

In the proposed approach, a CNN tackles the problem (5) because an explicit implementation of (5) represents a challenging task due to the multiple sources of noise. Indeed, mathematical approaches [45], [53] may fail to accurately model color representation. CNNs can process both the vial and color patches solving strongly nonlinear problems discerning relevant features inside images and can cope with the refraction and bubbles occurring inside the vial.

In practice, CNNs can learn (5) if trained properly with a dataset collecting images of the vial along with the reference patches. Our CNN processes a cropped image of the vial, together with a crop of the central part of all the color patches which are resized to the same size as the image of the vial, as shown in Fig. 4. The figure shows the proposed CNN meta-architecture. The network processes two inputs

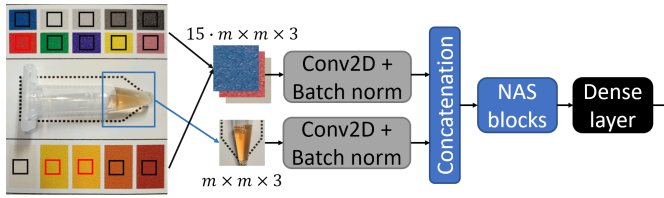


Fig. 4. CNN inputs are obtained by cropping the image. In the scheme of the CNN, NAS Blocks and Conv2D+ Batch norm refer to the part of the architecture obtained through the NAS procedure.

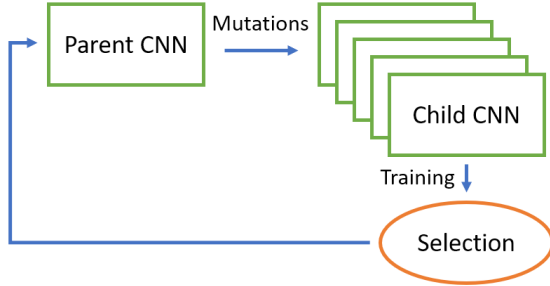


Fig. 5. Block scheme of the evolutionary algorithm used.

separately; each one passes through a convolutional layer and batch normalization layer. Then, a concatenation layer merges the information. The proposed structure prevents a strong bond between the more informative image of the vial and the images of color patches. The NAS procedure generates a single-branch architecture that processes the information from the two input branches. The network's output is the classification of the thiocyanate concentration level.

B. NAS With Input Selection

A blockwise NAS procedure selects the network architecture. Following a standard schema, three components characterized the procedure: the search space, the search algorithm, and the evaluation criteria.

1) *Search Space*: The search space S_s collects the candidate network architectures. In the present work, a candidate architecture is a stack of building blocks made of one convolutional layer and one batch normalization layer. The blocks embed three different parameters: the kernel size, the number of filters for each block, and the stride size. Together with the blocks' parameters, the search procedure tunes the number of blocks, the input size, and the color space used for the input images. Eventually, the combination of color patches is integrated into the network architecture to enhance the procedure.

2) *Evaluation Criteria*: The importance of each color reference is not known a priori. The NAS procedure measures it by generating networks that differ in weights assigned to the patches. The accuracy of the validation set drives the selection.

3) *Search Algorithm*: The search space is rather vast (or better, infinite). The evolutionary algorithm of Fig. 5 implements the exploration procedure. It generates a set of N "child" architectures starting from a "parent" network by applying random alterations to its structure. The alteration could change the set of reference patches. A parent yields seven different child networks by changing the number of blocks inside it, the three parameters describing the blocks, the color space used for the input images, their size, and which color patches are used. The child with the best evaluation score becomes the new

parent, and a new "offspring" will be generated from it. This search strategy allows a wide exploration of the search space with a simple implementation algorithm. Other strategies could tradeoff differently search time and quality of the solution [54], [55], however, a recent comparison suggested that the performance of the solution is similar for different approaches [56].

The procedure is summarized in Algorithm 1, with the following notation: the reference color space A_s contains all possible permutations of the reference colors A ; CNN_P indicates the parent architecture; $CNN_C(S_C, A_C) = R_m(CNN_P(S_P, A_P))$ denotes the child architecture generator function; N_c is the number of child architectures for each of the N_g generations; finally, $E(CNN, \mathcal{X}_V)$ is the evaluation function. After the initialization stage, the procedure generates the first parent architecture (line 1). For a preset number of iterations N_g , the approach generates and train child networks (lines 4 and 6, respectively). Then, the offspring are evaluated and the best one replaces the parent architecture (line 8).

Algorithm 1 Evolutionary NAS With Input Selection

Dataset Training set $\mathcal{X}_T = \{X_i, A_i, Y_i\}_{i=1, \dots, n}$ with X_i assay image, A_i reference color images, and Y_i the corresponding thiocyanate concentration class. Validation set $\mathcal{X}_V = \{X_i, A_i, Y_i\}_{i=1, \dots, m}$.

Pseudocode

- 1: **0. Init:** $CNN_P(S_P, A_P) = CNN_{P_0}(S_{P_0}, A_{P_0})$
 - 2: **for** g in N_g **do**
 - 3: **for** c in N_c **do**
 - 4: **1. Mutation:** $CNN_C(S_C, A_C) =$
 - 5: $R_m(CNN_P, A_P)$
 - 6: **2. Training:** Train $CNN_C(S_C, A_C)$ on \mathcal{X}_T
 - 7: **end for**
 - 8: **3. Selection:** $CNN_P(S_P, A_P) =$
 - 9: $\text{argmax}(E(\{CNN_C(S_C, A_C)\}_{j=1, \dots, N_c}, \mathcal{X}_V))$
 - 10: **end for**
-

IV. EXPERIMENTS

The section is organized as follows. Section IV-A presents the dataset and the experimental setup. Section IV-B reviews the experiments about the NAS procedure. Finally, Section IV-C compares the CNN architecture selected by NAS against two state-of-the-art methods developed to read semiquantitative and binary colorimetric assays performed on paper.

A. Experimental Setup

The experimental setup validates the robustness against moderate variation in lighting conditions when acquiring the results of the colorimetric assay using a smartphone in an indoor environment. The adopted protocol was inspired by the setup proposed in [57] and [23].

The images were collected in a laboratory with fixed lighting conditions (4000 K) without direct interaction with sunlight or other lighting sources. A lamp with three different light temperatures, corresponding to 3000, 4500, and 6000 K, was headed toward the vial changing its orientation and height

TABLE I

DATA SPLIT BETWEEN TRAINING, VALIDATION, AND TESTING SETS

WITH THE SMARTPHONE FLASH OFF AND ON		
Dataset	Flash off	Flash on
Training set	2.730	1.942
Validation set	1.226	956
Testing set	518	1.066
Total	4.474	3.964

to mimic real-world conditions. Different orientations induced shadow changes that distorted the color quantification process. Illumination intensities were measured using the light sensors equipped by the smartphone used to capture the pictures. The base illumination was 230 lux. The smallest illumination increment induced by the auxiliary lamp at the highest distance and the steepest inclination was 640 lux. The largest increment was 1725 lux. For each light condition and direction, a set of approximately 20 images was captured while moving the phone around to simulate an average user who could fail in aligning the smartphone with the test tube creating interaction with shadows produced by the vial.

Saliva from seven different healthy donors was collected over four weeks. To simulate a wide range of nonphysiological thiocyanate concentrations and to assess the smartphone-based semiquantitative detection performance, saliva samples were spiked with different biomarker concentrations, so to mimic ten levels of thiocyanate, covering the 0–2.50-mM range uniformly [3], [7], leading to a ten-class problem where each class corresponded to a different concentration level.

All images were captured by a Xiaomi 11 Lite 5G NE phone main camera. The experimental dataset contained 8.438 images, of which 4.474 were acquired with the flash switched off and 3.964 with the flash on. Table I shows the split between training, validation, and testing. Different sets of images included only images of vials with saliva acquired at different times and/or from different persons preventing possible overfitting on irrelevant saliva characteristics (e.g., the presence of bubbles or cloudiness inside the vial).

B. Definition of the CNN Architecture Through NAS Procedure

The initial parent architecture had a basic structure: the concatenation layer was directly linked with the dense layer. The input image size was 80×80 pixels, all the color patches shared the same weights, and the adopted color space was RGB. The kernel size, the stride size, and the number of filters for the first convolution were set to 1, 1, and 4, respectively. Each architecture was trained for 100 epochs with the Adam optimizer, categorical cross-entropy loss, and a learning rate of 10^{-3} using early stopping on the validation set with a patience value of 4 epochs. The number of generations N_g was 100. For each generation, only the child which presented the lowest validation loss was selected. Keras and TensorFlow Python libraries supported the NAS procedure implementation.

The architecture generated from the NAS procedure had an input size of 110×110 pixels in LAB color space and two blocks after the concatenation layer. Table II presents the architecture. Each row refers to network blocks composed of one convolutional layer and one batch normalization layer. The

TABLE II

NET ARCHITECTURE YIELDED FROM THE NAS PROCEDURE

Input	# Filters	Kernel size	Stride size
$110^2 \times 3 + 110^2 \times 36$	14	1×1	2×2
$55^2 \times 28$	18	1×1	3×3
$19^2 \times 18$	22	1×1	3×3

first column gives the input size for the corresponding block, while the other columns refer to, respectively, the number of filters, the kernel size, and the stride size of the convolutional layer inside the block. The first row of Table II shows the input of the network composed by the vial image, of size 110×110 pixels, and 12×3 channels dedicated to the color patches. The NAS procedure eventually selected 12 color patches out of 15; the discarded patches are marked with red squares in Fig. 4. This experiment confirmed the benefits of using these additional inputs to the neural network.

In total, the architecture involved 1.928 parameters and required 3.982.522 floating-point operations to process an image, making it suitable for deployment on a smartphone.

C. Comparison With the State of the Art

Recent surveys confirm the lack of works targeting the analysis of colorimetric assays in vials using a smartphone in an unconstrained scenario [20], [21]. In fact, the literature also lacks research targeting automatic readout for thiocyanate quantification. However, some recent works tackled similar problems. Two solutions for readout based on smartphone cameras were selected as the baseline. Both methods have been tuned to tackle the problem at hand. The methods were then trained and tested on the exact same set of data used for the proposed approach (as per Section IV-A).

The first baseline [23] is a CNN trained for automated readout of paper-based test strips for urine albumin. In this setup, the flash of the smartphone is exploited to bias the lighting setup, thus avoiding color references. The neural network for this method has been fed with a suitable cropped image of the vial resized to 64×64 pixel to enhance the prediction accuracy.

The second baseline [22] is a solution for smartphone-based colorimetric analysis of urine test strips for at-home prenatal care. The original approach uses an object detection network to identify the positions of the test strip and a few color reference pads. The test strip is compared with the reference color; the concentration value is obtained by computing the distance from the reference colors. Given that the focus of the present article is not on detection, the correct patches were manually extracted from the dataset; in other words, the setup considers the ideal case where all patches are correctly detected. The available patches in the left column of the custom card (Fig. 2) set the reference colors. Actually, the central (i.e., third) patch has been removed in this experiment, as empirical evidence suggested that the baseline improved its accuracy with such a setup. As a result, the experiment tackled a four-class classification problem.

The outcomes of the first experiment comparing the baseline [23] with the proposed work are presented in Table III. In this experiment, the flash was turned off. The table gives the

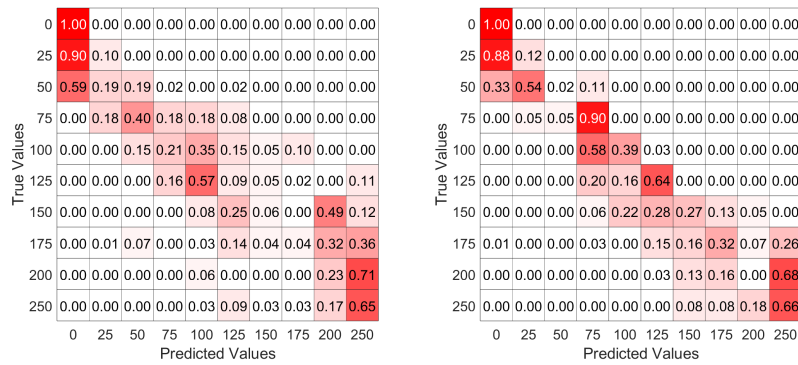


Fig. 6. Confusion matrix on the test set pictures captured with the flash off for the CNN of [23] (left) and the one presented in this article (right).

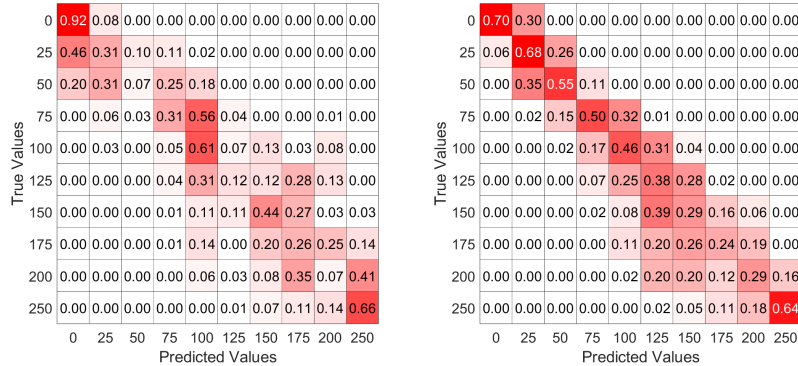


Fig. 7. Confusion matrix on the test set pictures captured with the flash on for the CNN of [23] (left) and the one presented in this article (right).

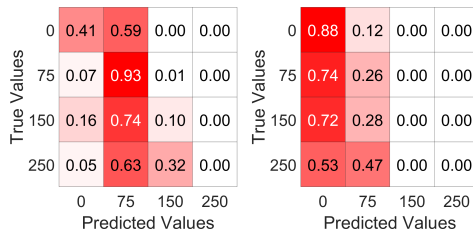


Fig. 8. Confusion matrix on the test set with the flash off (left) and on (right) using the hue distance [22] between the vial and reference color.

TABLE III

ACCURACY COMPARISON OF THE CNN PROPOSED IN [23] AND THE PROPOSAL, COMPUTED ON IMAGES CAPTURED WITH THE FLASH OFF

Method	Accuracy	Accuracy + 1
NAS	42.0%	80.9%
baseline	27.2%	65.2%

accuracy scored on the test set by, respectively, the CNN selected with the NAS procedure and the CNN proposed in [23]. The column Accuracy + 1 refers to the accuracy obtained when considering acceptable that the *predicted* class is at distance 1 from the *true* class. This metric is reasonable since the basal level of salivary thiocyanate has a certain level of intra- and interpersonal heterogeneity. Accordingly, different samples, belonging to different donors could be affected by a small bias in the original value of thiocyanate. Overall, experimental results show that the proposed approach outperforms the baseline.

Fig. 6 provides the confusion matrices characterizing such an experiment. The matrix on the left side refers to the baseline, while the matrix on the right side refers to the proposal. Each matrix gives all the possible combinations of

TABLE IV

ACCURACY COMPARISON OF THE CNN PROPOSED IN [23] AND THE PROPOSED CNNs, COMPUTED ON IMAGES CAPTURED WITH THE FLASH ON

Method	Accuracy	Accuracy + 1
NAS	46.8%	87.5%
baseline	38.1%	78.2%

true labels (rows) and the predicted labels (columns); for each combination, the normalized occurrences are provided. Each box is color-coded depending on its value with a color ranging from white, corresponding to 0, to pure red, corresponding to 1. The confusion matrices suggest that the proposed approach presented a higher accuracy, especially for elevated thiocyanate concentrations, where the assay color produces smaller changes in tone with respect to lower concentrations.

In the second experiment comparing the baseline [23] with the proposed work, the flash was turned on. Table IV presents the result; it has the same format as Table III. Both methods enhanced their accuracy with respect to the setup with the flash turned off, as suggested in [58] and [23] and the proposal outperformed again the baseline. Fig. 7 shows the confusion matrices obtained with this experiment.

Fig. 8 shows the outcomes of the experiment for the second baseline [22], which exploits as inputs the hue distances between the vial and four reference colors. The figure gives the confusion matrices obtained under two different setups: flash off (left) and flash on (right). In both cases, the baseline proved unsatisfactory, even with a four-class classification problem. In the setup with the flash off, the accuracy was about 39% and worsened with the other setup (about 32%).

V. CONCLUSION

The article presented a new method for automatic semiquantitative reading of thiocyanate concentration in saliva through assays performed in solution in ambient light. The approach consisted of a convolutional neural network, which exploits information on both the liquid sample and adjacent reference color images. The architecture for the networks was extracted using a NAS able to assert the importance of each reference color. The results confirm the validity of the proposed method, which leads to an acceptable level of accuracy.

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