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Noninvasive Assessment of Inspiratory Muscle Neuromechanical Coupling During Inspiratory Threshold Loading

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ABSTRACT Diaphragm neuromechanical coupling (NMC), which reflects the efficiency of conversion of neural activation to transdiaphragmatic pressure (Pdi), is increasingly recognized to be a useful clinical index of diaphragm function and respiratory mechanics in neuromuscular weakness and cardiorespiratory disease. However, the current gold standard assessment of diaphragm NMC requires invasive measurements of Pdi and crural diaphragm electromyography (oesEMGdi), which complicates the measurement of diaphragm NMC in clinical practice. This is the first study to compare invasive measurements of diaphragm NMC (iNMC) using the relationship between Pdi and oesEMGdi, with noninvasive assessment of NMC (nNMC) using surface mechanomyography (sMMG_{lic}) and electromyography (sEMG_{lic}) of lower chest wall inspiratory muscles. Both invasive and noninvasive measurements were recorded in twelve healthy adult subjects during an inspiratory threshold loading protocol. A linear relationship between noninvasive sMMG_{lic} and sEMG_{lic} measurements was found, resulting in little change in nNMC with increasing inspiratory load. By contrast, a curvilinear relationship between invasive Pdi and oesEMGdi measurements was observed, such that there was a progressive increase in iNMC with increasing inspiratory threshold load. Progressive recruitment of lower ribcage muscles, serving to enhance the mechanical advantage of the diaphragm, may explain the more linear relationship between sMMG_{lic} and sEMG_{lic} (both representing lower intercostal plus costal diaphragm activity) than between P_{di} and crural oesEMG_{di}. Noninvasive indices of NMC derived from sEMG_{lic} and sMMG_{lic} may prove to be useful indices of lower chest wall inspiratory muscle NMC, particularly in settings that do not have access to invasive measures of diaphragm function.

INDEX TERMS Electromyography, inspiratory threshold loading, mechanomyography, neuromechanical coupling, respiratory muscles.

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I. INTRODUCTION

Accurate assessment of respiratory muscle function is essential for both research and clinical practice [1]. Respiratory

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muscle function tests are particularly useful in the assessment of patients with respiratory symptoms [1] and for the diagnosis, phenotyping, and monitoring of patients with neuromuscular disease [2], [3]. The diaphragm is the principal muscle of inspiration [4], [5], and diaphragm weakness may present as breathlessness [6], [7], progressing to respiratory failure when weakness is severe [3]. Unlike electrical activation of the diaphragm, transdiaphragmatic pressure (Pdi) generation is dependent on neuromechanical coupling (NMC), i.e. the ability of the diaphragm muscle to transform electrical activation into pressure generation [8]. When the diaphragm fails to transform electrical activation into P_{di}, neuromechanical uncoupling occurs. Neuromechanical uncoupling is commonly observed in obstructive lung disease, such as chronic obstructive pulmonary disease, in which the force-generating capacity of the diaphragm is impaired by lung hyperinflation and altered chest wall geometry [9]. Neuromechanical uncoupling has been implicated in the perception of respiratory discomfort (breathlessness) in obstructive lung disease [10], [11] and in neuromuscular disease [12], [13], and is a factor determining successful weaning from mechanical ventilation [14]. Measurements of inspiratory muscle NMC could therefore provide clinically useful indices of inspiratory muscle function.

The gold standard assessment of human diaphragm pressure generation and neural respiratory drive requires invasive procedures, such as the balloon-catheter technique to measure P_{di} [1], and the use of a multipair esophageal electrode catheter to assess neural respiratory drive by quantifying the crural diaphragm electromyogram (oesEMG_{di}) [15]. These invasive tests can be uncomfortable for patients, require some skill from the operator involved, and are therefore rarely carried out in clinical practice. The development of novel, noninvasive indices of inspiratory NMC would therefore represent a significant advance in the assessment of patients with disordered respiratory mechanics.

The surface mechanomyogram (sMMG) is a noninvasive measure of muscle surface vibrations due to motor unit mechanical activity [16] and represents the mechanical counterpart of motor unit electrical activity as measured by surface electromyography (sEMG). Surface inspiratory muscle mechanomyographic [17]-[20] and electromyographic [19]-[21] signals can be recorded using lateral chest wall accelerometers and electrodes positioned over the lower intercostal spaces, proximal to the zone of apposition of the diaphragm (sMMG_{lic} and sEMG_{lic} respectively). These signals represent activation of the costal diaphragm [22], and of extradiaphragmatic chest wall and abdominal muscle recruitment [20], [23], [24] when respiratory effort increases [25], [26]. Using fixed sample entropy (fSampEn), an analysis technique which is less influenced by cardiac artefacts than conventional root mean square (RMS), we have nevertheless recently demonstrated strong correlations between Pdi and sMMGlic, and oesEMGdi and sEMGlic in healthy subjects during a submaximal inspiratory threshold loading protocol [20]. However, the use of sEMGlic and

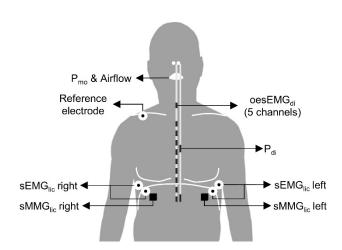


FIGURE 1. Sensor positioning for data acquisition.

sMMG_{lic} to assess NMC of the lower chest wall inspiratory muscles in a wholly noninvasive manner has not previously been investigated.

The principal aim of the study was, therefore, to investigate the combined use of sEMG $_{lic}$ and sMMG $_{lic}$ to calculate lower chest wall inspiratory muscle NMC in a wholly noninvasive manner, in comparison with diaphragm NMC calculated using the invasive oesEMG $_{di}$ and P_{di} measures, in healthy subjects during an incremental inspiratory threshold loading protocol.

II. METHODS

A. ETHICS STATEMENT

This study was approved by the NHS Health Research Authority (NRES Committee London – Dulwich 05/Q0703) and the experiments conformed to the standards of the Declaration of Helsinki. All subjects were fully informed of any risk associated with the study and provided their written consent before participation.

B. DATA ACQUISITION

Both invasive and non-invasive measurements (Fig. 1) of inspiratory muscle activation were recorded at the King's College London Respiratory Physiology Laboratory, King's College Hospital, London, United Kingdom. All signals were acquired from healthy adult subjects, with no history of cardiorespiratory or neuromuscular disease. Some of these signals have been analyzed in a previous study [20].

 P_{di} was calculated as the difference between gastric and esophageal pressures recorded using a dual-pressure transducer tipped catheter (CTO-2; Gaeltec Devices Ltd., Dunvegan, UK), as previously described [27], [28]. Crural oesEMG_{di} was recorded using a multipair esophageal electrode catheter (Yinghui Medical Equipment Technology Co. Ltd., Guangzhou, China) [15]. The pressure transducer and electrode catheters were inserted through the nose and optimally positioned as previously described [29], [30]. sEMG_{lic} was recorded bilaterally using two pairs of disposable surface Ag/AgCl electrodes (H124SG; Covidien Kendall) placed on



the skin over the seventh or eighth intercostal spaces, between the mid-axillary and the anterior axillary lines [19]–[21]. sMMG $_{\rm lic}$ was recorded using two triaxial accelerometers (TSD109C2; BIOPAC Systems Inc, CA, USA) attached bilaterally to the skin with adhesive rings as close as possible to the sEMG $_{\rm lic}$ electrodes along the seventh or eighth intercostal space, over the anterior axillary line [17], [20]. Respiratory flow was measured using a pneumotachograph (4830; Hans Rudolph Inc, KS, USA) connected to a differential pressure transducer (DP45; Validyne Engineering, CA, USA). Mouth pressure ($P_{\rm mo}$) was measured from a side port on the pneumotachograph using a second differential pressure transducer (MP45; Validyne Engineering, CA, USA).

The oesEMG_{di} and sEMG_{lic} signals were amplified (gain 100), high-pass filtered at 10 Hz, and AC-coupled before acquisition (CED 1902; Cambridge Electronic Design Limited, Cambridge, UK). All signals were acquired using a 16-bit analogue-to-digital converter (PowerLab 16/35; ADInstruments Ltd, Oxford, UK) and displayed on a laptop computer running LabChart software (Version 7.2, ADInstruments Pty, Colorado Springs, USA) with analogue to digital sampling at 100 Hz (flow and pressures), 2000 Hz (sMMG_{lic}), and 4000 Hz (oesEMG_{di} and sEMG_{lic}).

C. PROTOCOL

1) MAXIMAL VOLITIONAL MANEUVERS

Two maximal volitional inspiratory maneuvers were performed initially: maximal inspiratory effort against an occluded mouthpiece (a Mueller maneuver) from functional residual capacity to determine maximal inspiratory mouth pressure (PImax) [1], and maximal inspiration to total lung capacity [30], [31]. These maneuvers were performed sitting upright in a chair with a noseclip in place and were repeated several times to ensure maximal volitional effort. All measurements were recorded continuously during the maneuvers. Each participant's PImax was used to determine the inspiratory threshold loads used in their individual incremental inspiratory threshold loading protocol.

2) INSPIRATORY THRESHOLD LOADING PROTOCOL

All participants performed an inspiratory threshold loading protocol at five inspiratory threshold loads set at 12% (L_1), 24% (L_2), 36% (L_3), 48% (L_4), and 60% (L_5) of the subject's PImax. Inspiratory threshold loads were generated using an electronic inspiratory muscle trainer (POWERbreathe K5; POWERbreathe International Ltd, Southam, UK) attached to the distal end of the pneumotachograph. Subjects were seated and breathed through the pneumotachograph via a mouthpiece with a noseclip in place. Baseline measurements were recorded during a minimum of 2 minutes of quiet tidal breathing (L_0), following which the inspiratory muscle trainer was attached to the pneumotachograph and the series of threshold loads was imposed. Subjects were not provided with any specific instructions to adopt a certain duty cycle and were free to choose their own breathing frequency. Subjects were,

however, informed that effort was needed to overcome the threshold loads, and they were therefore encouraged to focus on using their diaphragm, to perform quick deep inspirations and to ensure that expiration was complete before making their next inspiratory effort. Each load consisted of 30 breaths at most followed by a resting period to allow all respiratory measures to return to baseline.

D. DATA ANALYSIS

1) DATA PRE-PROCESSING

LabChart data were exported as Matlab files, and analyzed offline in MATLAB (The MathWorks, Inc., vR2014a, Natick, MA, USA).

oesEMG_{di} and sEMG_{lic} signals were resampled at 2000 Hz and filtered with an 8th-order zero-phase Butterworth band-pass filter between 10 and 600 Hz, and with a 2-Hz bandwidth notching comb filter to remove the power line interference at 50 Hz and all its harmonics. sMMG_{lic} signals were resampled at 500 Hz and filtered with an 8th-order zero-phase Butterworth band-pass filter between 5 and 40 Hz. After filtering, the total acceleration measured by each accelerometer (|sMMG_{lic}|) was arithmetically calculated as the norm of the vector formed by its three sMMG_{lic} components (sMMG_{lic} X, sMMG_{lic} Y, and sMMG_{lic} Z). P_{mo} and PImax were expressed as absolute values.

All signals were segmented into inspiratory and expiratory signal segments by means of a zero-crossing detector on the P_{mo} signal. After segmentation, all signal segments were visually inspected and those either containing artefacts within the EMG and MMG signals or having an unusual pressure pattern were rejected. In general, less than 30% of respiratory cycles per load were rejected, and these typically occurred at the beginning and the end of each load. The following parameters were then calculated for each respiratory cycle: inspiratory time, duty cycle (inspiratory time / total breathing cycle time), breathing frequency, and inspiratory volume. Inspiratory volume was calculated as the area under the curve of the inspiratory flow trace. The median values of all respiratory cycles during resting breathing and threshold loading were then calculated and ten cycles that contained the four parameters nearest to the median values were automatically selected, resulting in a total of sixty respiratory cycles for each subject.

2) MYOGRAPHIC SIGNAL ANALYSIS USING fSampEn

Sample entropy is a measure of regularity and complexity of time-series signals [32], so that more regular signals are less complex and lead to lower values of sample entropy. The following parameters must be fixed in sample entropy: the embedded dimension, m, and the tolerance, r. The latter is usually set as a percentage of the standard deviation of the signal analyzed. In fSampEn, however, sample entropy is calculated within a moving window, instead of over a whole signal, using a fixed r value for all windows. In this way,



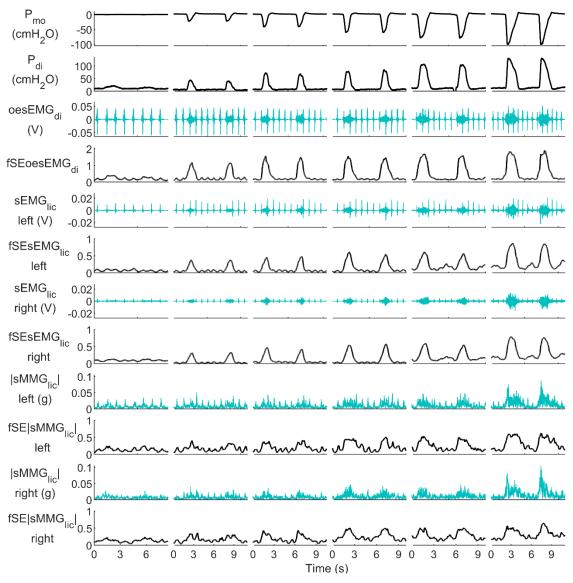


FIGURE 2. Signals recorded during the inspiratory threshold loading protocol in a single healthy subject. Two respiratory cycles are shown during quiet resting breathing, and during inspiratory threshold loading at 12%, 24%, 36%, 48%, and 60% of Plmax. The oesEMG_{di} signal shown is the signal recorded from electrode pair 1.

fSampEn is not only sensitive to changes in signal complexity, but also to changes in signal amplitude [33].

In myographic respiratory signals, fSampEn has proven to be less sensitive in quantifying amplitude variations of more deterministic signal components, such as cardiac artefacts, than in quantifying amplitude variations of more complex signal components, such as inspiratory muscle EMG and MMG [21], [33].

In this study, fSampEn signals were calculated for the oesEMG_{di} (fSEoesEMG_{di}), sEMG_{lic} (fSEsEMG_{lic}), and $|sMMG_{lic}|$ (fSE $|sMMG_{lic}|$) signals (Fig. 2), using a moving window of 500 ms with a 50-ms step, m equal to 2 and r equal to 0.05 (fSEoesEMG_{di}), 0.3 (fSEsEMG_{lic}), and 0.5 (fSE $|sMMG_{lic}|$) times the global standard deviation of each signal. These are the optimal general fSampEn parameters

for inspiratory muscle activity estimation as we previously described [34].

The level of inspiratory muscle activity during each respiratory cycle was calculated as the mean inspiratory P_{di} , fSEoesEMG_{di}, fSEsEMG_{lic}, and fSE|sMMG_{lic}| signals. For fSEoesEMG_{di}, the highest mean value obtained across all five bipolar electrode pairs was selected. Mean P_{di} was calculated after removal of the baseline from the P_{di} signal, which was determined as the moving minimum using a moving window of 1.5 times the maximum inspiratory time of each load.

The per-breath mean values of P_{di} , fSEoesEMG_{di}, fSEsEMG_{lic}, and fSE|sMMG_{lic}| signals were expressed as percentages of their respective largest mean values obtained throughout the inspiratory threshold loading protocol and the two maximal volitional



maneuvers (P_{di%max}, fSEoesEMG_{di%max}, fSEsEMG_{lic%max}, and fSE|sMMG_{lic}|_{% max}).

3) INSPIRATORY MUSCLE NMC

Inspiratory muscle NMC was calculated as the ratio of a mechanical measure ($P_{di\%max}$ or $fSE|sMMG_{lic}|_{\% max}$) to an electrical measure (fSEoesEMG $_{di\%max}$ or fSEsEMG $_{lic\%max}$). Depending on the measures involved, an invasive NMC (iNMC), mixed NMC (mNMC), or noninvasive NMC (nNMC) was calculated as in (1), (2), and (3), respectively.

$$iNMC_{L_i} = \frac{\overline{P_{di\% \max - L_i}}}{\overline{fSEoesEMG_{di\% \max - L_i}}}$$
(1)

$$mNMC_{L_i} = \frac{P_{di\% \max - L_i}}{\overline{fSEsEMG_{lic\% \max - L_i}}}$$
 (2)

$$iNMC_{L_{i}} = \frac{\overline{P_{di\% \max - L_{i}}}}{\overline{fSEoesEMG_{di\% \max - L_{i}}}}$$

$$mNMC_{L_{i}} = \frac{\overline{P_{di\% \max - L_{i}}}}{\overline{fSEsEMG_{lic\% \max - L_{i}}}}$$

$$nNMC_{L_{i}} = \frac{\overline{fSE \mid sMMG_{lic} \mid_{\% \max - L_{i}}}}{\overline{fSEsEMG_{lic\% \max - L_{i}}}}$$

$$(3)$$

where i goes from 0 to 5, and the overlines represent the average value of the ten respiratory cycles selected for resting breathing and each load.

E. STATISTICAL ANALYSIS

Normality of all measures and parameters calculated for each subject was tested using Lilliefors tests. Since not all measures and parameters had a normal distribution, all data are expressed as median and interquartile range.

Changes in inspiratory and expiratory times, duty cycle, respiratory frequency, and inspiratory volume during threshold loading were assessed using Friedman tests and multiple Wilcoxon signed-rank tests with Bonferroni adjusted

Linear relationships between measures of inspiratory muscle activity could not be assumed a priori, and therefore the relationships between electrical and mechanical measures during threshold loading were assessed individually using Spearman's rank correlation coefficients (ρ).

Changes in iNMC, mNMCs, and nNMCs during threshold loads L1-L5 were firstly assessed for the 12 subjects using Friedman tests, followed by multiple Wilcoxon signed-rank tests with Bonferroni adjustment for multiple comparisons. Secondly, linear regression analysis was used individually to assess whether iNMC, mNMCs, and nNMCs increased significantly during threshold loads L1-L5.

The significance level for all statistical tests was set at 0.05.

III. RESULTS

Twelve subjects (6 male and 6 female, age 33 (30-39) years, body mass index 22.2 (20.6-24.2) kg/m², forced expiratory volume in 1 second 98.0 (94.8-105.5) % of predicted, forced vital capacity 105.0 (91.5-110.2) % of predicted, forced expiratory volume in 1 second/forced vital capacity 81.9 (74.1-83.9) %) were included in the study and completed the incremental inspiratory loading protocol.

A. BREATHING PATTERN AND PRESSURE GENERATION

The group median (interquartile range) PImax was 87.0 (78.0-116.5) cmH₂O. The inspiratory threshold loads increased from 11 (10-14) cmH₂O during L1 to 52 (47-70) cmH₂O during L5. P_{mo} decreased from 0.3 (0.3-0.4) cmH_2O during L0 to 53.7 (49.5-68.4) cmH_2O (p < 0.001) during L5. P_{di} increased from 11.1 (7.1-11.7) cmH₂O at rest to $64.4~(46.1-67.9)~cmH_2O~(p < 0.001)$ at L5. Evolution of inspiratory time, duty cycle, respiratory frequency, and inspiratory volume during threshold loading is shown in Fig. 3. No significant differences were found in inspiratory time, duty cycle, and respiratory frequency during the inspiratory threshold loading protocol. Significant differences were found in expiratory time between resting breathing and the highest loads L4 and L5. Significant differences were also found in inspiratory volume between resting breathing and all threshold loads, and between load L1 and loads L3-L5. These results indicate that the breathing strategies adopted during successive stages of the inspiratory threshold loading protocol were similar.

B. INVASIVE VS NONINVASIVE NMC

Fig. 4a-e show the relationships between electrical and mechanical measures of inspiratory muscle activity, measured both invasively (fSEoesEMGdi%max and Pdi%max respectively) and noninvasively (fSEsEMG_{lic%max} and fSE|sMMG_{lic}|_{% max} respectively). Table 1 shows strong to very-strong correlations between fSEoesEMG $_{di\%max}$ and P_{di%max}, fSEsEMG_{lic%max} and P_{di%max}, and fSEsEMG_{lic%max} and $fSE|sMMG_{lic}|_{\% max}$.

The relationship between fSEoesEMGdiimax and Pdiimax was more curvilinear than the relationships between fSEsEMG_{lic%max} and either P_{di%max} or fSE|sMMG_{lic}|_{% max}. As explained in Appendix A, the relationship between fSEoesEMG_{di%max} and P_{di%max} is well described by an exponential model. Accordingly, iNMC (Fig. 4f) increased progressively and significantly from load L1 to load L5. In contrast, the relationships between fSEsEMG_{lic%max} and either $P_{di\%max}$ or $fSE|sMMG_{lic}|_{\% max}$ are well described by linear models, so that mNMCs (Fig. 4g and 4h) and nNMCs (Fig. 4i and 4j) increased very little, non-significantly, from load L1 to load L5. Table 2 shows individual linear regression results for iNMC, mNMC, and nNMC against inspiratory loads L1-L5. Statistically significant positive relationships were found between iNMC and inspiratory load (% PImax) in all subjects, in contrast to the absence of a significant relationship between mNMC or nNMC and inspiratory load in the majority of individuals studied.

IV. DISCUSSION AND CONCLUSION

The current gold standard assessment of diaphragm NMC in human subjects requires invasive measurements of Pdi and oesEMG_{di}. These measurements are technically difficult and often uncomfortable for the individual being studied, presenting a barrier to the measurement of diaphragm



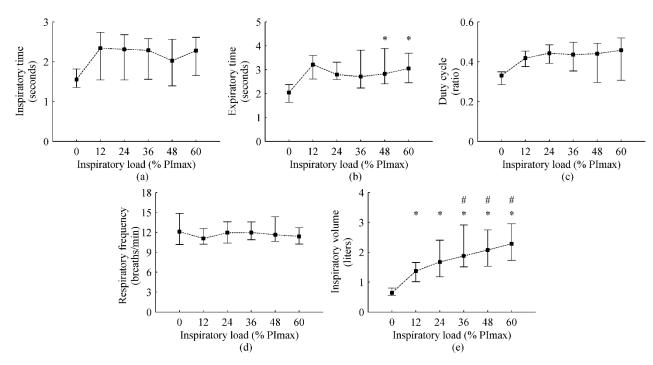


FIGURE 3. Evolution of inspiratory time, duty cycle, respiratory frequency, and inspiratory volume during progressive inspiratory threshold loading. Data points represent median and interquartile range of the 12 subjects for each load. Dashed black lines show the order of execution of the inspiratory threshold loads. Symbols * and # indicate statistically significant differences with respect to L0 (0% Plmax) and L1 (12% Plmax), respectively.

TABLE 1. Spearman's ρ between electrical and mechanical measures of inspiratory muscle activity.

Subject ID	fSEoesEMG _{di%max} - P _{di%max}	fSEsEMG _{lic%max} left- P _{di%max}	$\begin{array}{c} \text{fSEsEMG}_{\text{lic}\%\text{max}} \text{ right-} \\ P_{\text{di}\%\text{max}} \end{array}$	fSEsEMG _{lic%max} left- fSE sMMG _{lic} _{%max} left	fSEsEMG _{lic%max} right- fSE sMMG _{lic} _{%max} right	
1	0.85	0.95	0.92	0.81	0.89	
2	0.94	0.94	0.93	0.84	0.88	
3	0.80	0.82	0.84	0.94	0.91	
4	0.77	0.90	0.93	0.90	0.94	
5	0.90	0.96	0.89	0.94	0.89	
6	0.80	0.84	0.84	0.89	0.84	
7	0.76	0.96	0.96	0.94	0.96	
8	0.71	0.97	0.97	0.95	0.94	
9	0.93	0.96	0.96	0.93	0.93	
10	0.64	0.78	0.81	0.89	0.83	
11	0.57	0.73	0.76	0.79	0.76	
12	0.89	0.88	0.86	0.87	0.91	
Median (IQR)	0.80 (0.75-0.89)	0.92 (0.84-0.96)	0.90 (0.84-0.94)	0.90 (0.86-0.94)	0.90 (0.87-0.93)	

NMC outside specialist research and clinical settings. This is the first study to compare measurements of diaphragm NMC using invasive techniques (the relationship between P_{di} and oesEMG_{di}) with noninvasive assessment of lower chest wall inspiratory muscle NMC using sMMG_{lic} and sEMG_{lic} measurements. Wholly noninvasive indices of lower chest wall inspiratory muscle NMC (nNMC, i.e., the ratio of fSE|sMMG_{lic}| $_{\%$ max to fSEsEMG_{lic $_{\%}$ max}) exhibited an immediate decrease at the onset of inspiratory loading (the transition from resting breathing to 12% PImax), followed by little and nonsignificant changes over progressive increases in inspiratory load between 12% and 60% of PImax. This is due to the mostly linear increase in fSE|sMMG_{lic}| $_{\%}$ max

relative to fSEsEMG $_{lic\%max}$ at increasing inspiratory loads. There was also a linear relationship between noninvasive fSEsEMG $_{lic\%max}$ and invasive $P_{di\%max}$ measurements, again resulting in little change in mNMC with increasing inspiratory load. By contrast, the relationship between invasive fSEoesEMG $_{di\%max}$ and $P_{di\%max}$ measurements was observed to be curvilinear, such that there was a progressive increase in iNMC with progressive increases in inspiratory threshold load

Before considering these differences further, it is pertinent to review the key steps in the transformation of neural activation of the diaphragm, quantified as $oesEMG_{di}$, into P_{di} . The first step is translation of $oesEMG_{di}$ to muscle force, and

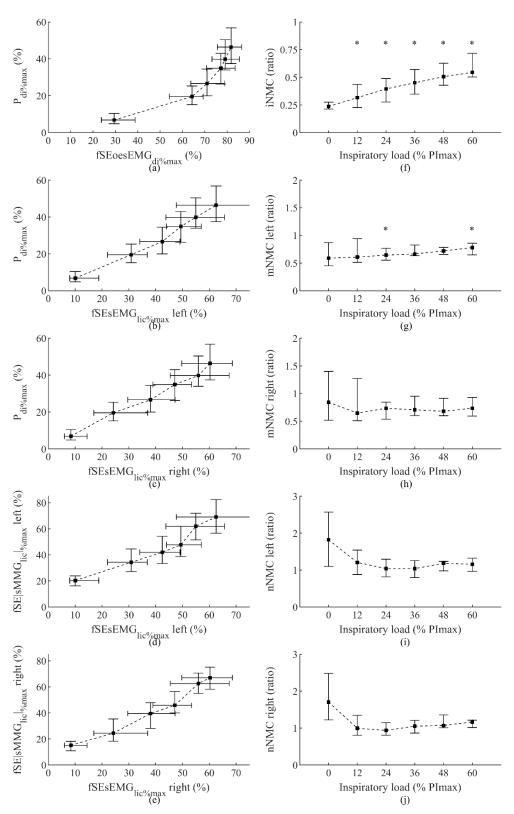


FIGURE 4. Relationship between electrical and mechanical measures of inspiratory muscle activation, and the corresponding NMC ratios, recorded during the incremental inspiratory threshold loading protocol. Invasive (iNMC), mixed (mNMC), and noninvasive (nNMC) NMCs were calculated as the ratios of $P_{di\%max}$ to fSEoesEMG $_{di\%max}$, $P_{di\%max}$ to fSEsEMG $_{lic\%max}$, and fSE $_{lic\%max}$ and fSE $_{lic\%max}$ to fSEsEMG $_{lic\%max}$, respectively. Data points represent median and interquartile range of the 12 subjects for each load. Dashed black lines show the order of execution of the inspiratory threshold loads. All data points with a * symbol were significantly different to each other.



TABLE 2. Linear regression between NMCs and inspiratory loads L1-L5.

Subject	iNMC		mNMC left		mNMC right		nNMC left		nNMC right	
ID	Slope	p value	Slope	p value	Slope	p value	Slope	p value	Slope	p value
1	0.067	0.002*	0.058	0.001*	0.052	0.077	0.040	0.603	0.060	0.185
2	0.063	0.027*	0.043	0.081	-0.061	0.273	0.004	0.917	0.126	0.175
3	0.057	0.043*	-0.119	0.212	-0.141	0.226	0.026	0.685	-0.098	0.160
4	0.053	0.024*	-0.003	0.892	-0.014	0.581	0.065	0.214	0.120	0.048*
5	0.060	0.010*	0.029	0.013*	0.004	0.095	0.024	0.301	0.138	0.032*
6	0.088	0.001*	0.017	0.734	-0.068	0.441	0.016	0.797	0.036	0.500
7	0.075	0.008*	-0.027	0.562	-0.024	0.501	-0.139	0.102	-0.046	0.018*
8	0.087	0.000*	0.041	0.144	0.035	0.043*	-0.043	0.192	-0.036	0.247
9	0.045	0.009*	0.045	0.053	-0.007	0.773	-0.117	0.236	-0.113	0.251
10	0.077	0.027*	0.041	0.373	0.029	0.500	0.039	0.330	0.037	0.389
11	0.063	0.001*	0.052	0.064	0.052	0.011*	0.019	0.784	0.050	0.099
12	0.046	0.012*	0.055	0.059	-0.003	0.949	-0.046	0.479	0.004	0.960

*Significant p values (<0.05)

the second is translation of diaphragm muscle force to P_{di}. As in other striated skeletal muscle, the relationship between oesEMG_{di} and diaphragm force of contraction depends on the force-length, force-velocity, and force-frequency responses of the muscle [35]. Most or all of the reduction in P_{di} after increasing lung volume can be explained by the force-length properties of the diaphragm [36]. The relationship between diaphragm force and P_{di} depends on diaphragm curvature and chest wall (thoracoabdominal) configuration [37]. Grassino et al. observed that the slope of the relationship between crural oesEMGdi and Pdi during isometric diaphragmatic contractions at increasing lung volumes became steeper as ribcage and abdominal anteroposterior (AP) diameters decreased, indicating that the principal determinant of the gain in P_{di} relative to neural activation (i.e. the mechanical advantage of the diaphragm) at a given lung volume is thoracoabdominal configuration [37].

Observations of curvilinear relationships between fSEoesEMG_{di%max} and P_{di%max} in the present study are in keeping with previous work [38], [39]. Laghi et al. [39] attributed the apparent increased mechanical advantage of the diaphragm during inspiratory threshold loading to progressive increases in extradiaphragmatic muscle contribution to tidal breathing, expiratory muscle recruitment, and decreased end-expiratory lung volume. Expiratory muscle recruitment during inspiratory threshold loading decreases abdominal-wall compliance [40], and therefore may facilitate inspiration by increasing diaphragmatic length [41]. Decreased abdominal compliance can also increase the fulcrum effect of the abdominal contents on the diaphragm, thereby enhancing displacement/elevation of the lower ribcage by diaphragm contraction during inspiration [42]. Expiratory muscle recruitment has also been associated with progressive reductions in end-expiratory lung volume during inspiratory threshold loading, with consequent improvement in the mechanical advantage of the diaphragm [37], [38], [40], [43], [44]. Indeed, under conditions of increased ventilatory demand, increased mechanical output of the ribcage and abdominal muscles serves to optimize the mechanical advantage of the diaphragm by permitting the diaphragm to operate at a greater overall length and limiting velocity of shortening [45].

In contrast to the curvilinear relationship between P_{di%max} and fSEoesEMG_{di%max}, relationships between $P_{di\%max}$ and $fSEsEMG_{lic\%max}$, and $fSE|sMMG_{lic}|_{\%max}$ and fSEsEMG_{lic%max} were linear. Whereas oesEMG_{di} signals are specific to the crural diaphragm [46], sEMGlic and sMMG_{lic} signals recorded over the lower chest wall capture activation of the costal portion of the diaphragm [22] and are vulnerable to cross talk from extradiaphragmatic chest wall and abdominal muscle activation [20], [23], [24]. Although the diaphragm is the principal respiratory muscle engaged at rest [47], during increased ventilation [47] or inspiratory loading [48], [49], extradiaphragmatic muscle activity increases to allow the diaphragm to function under optimal length-tension characteristics [45]. Therefore, it is possible that the observation of a more linear relationship between P_{di%max} and fSEsEMG_{lic%max}, and between fSE|sMMG_{lic}|_{% max} and fSEsEMG_{lic}_{%max}, than between P_{di%max} and fSEoesEMG_{di%max}, results, in part, from progressive increases in extradiaphragmatic muscle activation serving to optimize the mechanical advantage of the diaphragm as inspiratory load increased. Crosstalk from extradiaphragmatic muscles would be expected to contribute to the sEMGlic and sMMGlic signals (and therefore to nNMC and mNMC), but not to oesEMGdi or iNMC because the latter measures are specific to the diaphragm. An apparent "left-shift" in the relationship between P_{di%max} and fSEsEMG_{lic%max}, when compared to the relationship between P_{di%max} and fSEoesEMG_{di%max} (Fig. 4), may also reflect greater neuromechanical efficiency of the costal vs crural diaphragm as recently demonstrated in a canine model [50].

An important advantage of this study is the use of fSampEn to analyze oesEMG_{di}, sEMG_{lic} and $|sMMG_{lic}|$ signals. These signals are highly contaminated by cardiac artefacts, which complicates the analysis especially using conventional RMS as in previous studies [19], [27], [29]–[31], [38], [39], [51]. Thus, an RMS-based analysis of myographic signals requires signal conditioning to reduce or eliminate cardiac



TABLE 3. Parameter estimates of the linear and exponential regression models to describe the relationships between electrical and mechanical measures
of inspiratory muscle activity.

	Linear model				Exponential model					
	a_{l}	$\operatorname{CI} a_l$	$b_{\it l}$	CI $b_{\it l}$	\mathbb{R}^2	a_e	CI $a_{\it e}$	$b_{\it e}$	CI $b_{\it e}$	\mathbb{R}^2
E ₁ =fSEoesEMG _{di%max} M ₁ =P _{di%max}	0.68	(0.28, 1.08)	-16.75	(-44.40, 10.91)	0.85	1.03	(0.09, 1.96)	0.046	(0.034, 0.058)	0.99
E_2 =fSEsEMG _{lic%max} left M_2 = $P_{di\%max}$	0.76	(0.62, 0.89)	-2.53	(-8.60, 3.54)	0.98	7.48	(4.29, 10.66)	0.030	(0.022, 0.038)	0.98
E_3 =fSEsEM $G_{lic\%max}$ right M_3 = $P_{di\%max}$	0.72	(0.62, 0.83)	0.74	(-3.64, 5.12)	0.99	8.98	(4.71, 13.24)	0.027	(0.018, 0.036)	0.97
E ₄ =fSEsEMG _{lic%max} left M ₄ =fSE sMMG _{lic} _{%max} left	0.92	(0.60, 1.24)	7.59	(-6.83, 22.02)	0.94	15.81	(11.32, 20.29)	0.024	(0.018, 0.029)	0.98
$\begin{array}{l} E_5 \!\!=\!\! fSEsEMG_{lic\%max} \\ right \\ M_5 \!\!=\!\! fSE sMMG_{lic} _{\%max} \\ right \end{array}$	1.02	(0.76, 1.28)	2.71	(-8.46, 13.88)	0.97	12.83	(9.85, 15.81)	0.028	(0.023, 0.032)	0.99

artefacts. However, fSampEn allows myographic signals to be analyzed with less interference from cardiac artefacts and without the need for prior rejection of cardiac artefacts. A combined analysis of sEMG_{lic} and |sMMG_{lic}| using fSampEn would therefore allow NMC to be calculated noninvasively with minimal contamination by cardiac noise. Normalizing fSampEn of oesEMG_{di}, sEMG_{lic} and |sMMG_{lic}| signals to that evoked during maximal volitional maneuvers provides measurements of the activity of inspiratory muscles relative to their capacity which are comparable between subjects. fSampEn-based measurements from inspiratory muscle myographic signals strongly correlate with the corresponding RMS-based measurements from the same signals, as described in previous studies [20], [21], [33]. Although both fSampEn and RMS can be used to analyze the amplitude of a signal, it is important to note, however, that fSampEn and RMS are conceptually different and, in fact, do not have the same units. Like RMS, fSampEn can track amplitude variations of a signal, and therefore can be used to estimate inspiratory muscle activation from myographic signals [21], [33]. However, fSampEn does not only depend on the amplitude, but also on the complexity of a signal. Therefore, fSampEn- and RMS-based measurements are not interchangeable.

The noninvasive nature of the sEMG_{lic} and sMMG_{lic} measurements is of potential value to clinical assessment of patients with cardiorespiratory and neuromuscular disease, as well as to future research studies of respiratory muscle function. The most powerful prognostic biomarker for mortality stratification in amyotrophic lateral sclerosis is twitch trans-diaphragmatic pressure [3]. The value of noninvasive indices of neuromechanical activation in patients with progressive neuromuscular disease is therefore worthy of further study. Increased neural respiratory drive, consequent to increased inspiratory muscle loading and neuromechanical uncoupling of the diaphragm, is strongly implicated in the perception of breathlessness and exercise limitation in

chronic respiratory disease [10], [52]–[55]. The development of noninvasive objective indices of NMC of the respiratory muscles would therefore be of value both to the clinical assessment of breathlessness patients, and mechanistic investigations of interventions that reduce breathlessness and increase exercise tolerance [56], [57].

We acknowledge the limitations of our study, which present opportunities for future investigation. Our study cohort was relatively small, comprising of twelve healthy subjects. The sample size reflects the challenges associated with recruitment to studies using invasive measures of diaphragm function. The potential clinical utility of noninvasive NMC measurements should be tested in disease populations such as COPD. The body mass index values of our participants were within the normal range. Since thicker layers of body fat act as attenuator filters for sMMG [58] and sEMG [59] signals, the effect of body composition and body mass index on the relationships between sMMG_{lic} and sEMG_{lic} measures in otherwise healthy individuals should be a focus of future research.

In conclusion, we have used 7th/8th intercostal space sEMG and sMMG recordings to derive noninvasive indices of lower chest wall inspiratory muscle NMC in healthy adults during an incremental inspiratory threshold loading protocol. In contrast to the significant increases in iNMC, derived as the ratio of P_{di%max} to fSEoesEMG_{di%max}, there were little and nonsignificant changes in mNMC and nNMC during incremental inspiratory threshold loading, since the relationships between fSE|sMMG $_{lic}$ |% max and fSEsEMG $_{lic}$ %max, and P_{di%max} and fSEsEMG_{lic%max} were linear. We postulate that these differences may reflect increasing crosstalk from extradiaphragmatic lower chest wall and abdominal muscle activation to sEMGlic and sMMGlic signals (and hence to mNMC and nNMC), but not to crural oesEMG_{di} signals and iNMC, the latter measures being specific to the diaphragm. Noninvasive indices of NMC derived from sEMGlic and



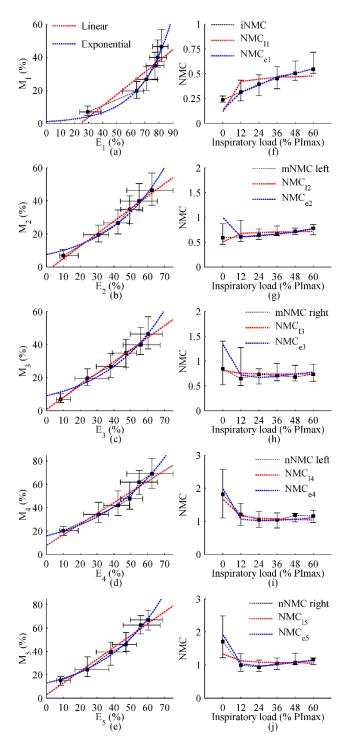


FIGURE 5. Linear (red) and exponential (blue) models fitted for the relationships between electrical and mechanical measures of inspiratory muscle activity. Black squares and bars represent median and interquartile range of the 12 subjects for each load.

sMMGlic may prove to be useful indices of lower chest wall inspiratory muscle NMC, particularly in settings that do not have access to invasive measures of diaphragm function.

APPENDIX

THEORETICAL EXPLANATION FOR THE EVOLUTION OF NMC DURING INSPIRATORY THRESHOLD LOADING

Inspiratory muscle NMC was calculated as the ratio of a mechanical measure to an electrical measure, and therefore the evolution of NMC during inspiratory threshold loading depends on the relationship between the measures involved in its calculation. The relationship between an electrical (E_i) and a mechanical (M_i) measure can be described in the simplest way by means of either a linear or an exponential function as in (A1) and (A2), respectively.

$$M_i = a_l \times E_i + b_l \tag{A1}$$

$$M_i = a_e \times e^{b_e \times E_i} \tag{A2}$$

where a_l, b_l, a_e , and b_e are the parameters of the models. The NMC, therefore, at a given level of inspiratory muscle electrical and mechanical activation can be expressed as in (A3) (linear case) and (A4) (exponential case).

$$EMC_{l} = \frac{M_{i}}{E_{i}} = a_{l} + \frac{b_{l}}{E_{i}}$$

$$EMC_{e} = \frac{M_{i}}{E_{i}} = \frac{a_{e} \times e^{b_{e} \times E_{i}}}{E_{i}}$$
(A3)

$$EMC_e = \frac{M_i}{E_i} = \frac{a_e \times e^{b_e \times E_i}}{E_i} \tag{A4}$$

According to (A3) and (A4), NMC₁ and NMC_e values tend to be more dispersed for low values of E_i. However, NMC_l tends to a constant value (a_l) as E_i increases, whereas NMC_e increases.

Parameters of linear and exponential models were estimated using the median values of fSEoesEMGdi%max and $P_{di\%max}$, $fSEsEMG_{lic\%max}$ and $P_{di\%max}$, and $fSEsEMG_{lic\%max}$ and fSE|sMMG_{lic}|% max, of the 12 subjects for each load (see Table 3). Adjusted curves are shown together with data points in Fig. 5a-e. The relationship between fSEoesEMG_{di%max} and P_{di%max} was better described by the exponential model ($R^2 = 0.99$) than the linear model ($R^2 =$ 0.85). However, the relationships between fSEsEMG_{lic%max} and either P_{di%max} or fSE|sMMG_{lic}|_{% max} were well described by both linear (R² ranging from 0.94 to 0.99) and exponential (R² ranging from 0.97 to 0.99) models, so the simplest model was selected, which was the linear model.

Theoretical NMCs were calculated using the estimated coefficients of linear and exponential models, as in (A3) and (A4). Fig. 5f-j show theoretical NMCs together with NMCs calculated using the acquired data. iNMC clearly increased with increasing load, so that it is better adjusted by the theoretical NMC derived from an exponential model. mNMC and nNMC, however, remained almost constant or increased slightly during threshold loads L1-L5, indicating that the relationships between fSEsEMGlic%max and either P_{di%max} or fSE|sMMG_{lic}|_{% max} can be well explained by a linear model.

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